# International Journal of Livestock Production

Volume 8 Number 1 January 2017 ISSN 2141-2448



## **ABOUT IJLP**

The International Journal of Livestock Production (IJLP) is an open access journal that provides rapid publication (monthly) of articles in all areas of the subject such as Selective breeding in animal husbandry, the health effects of animal cruelty, fishery in terms of ecosystem health, Fisheries acoustics etc.

The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in the IJLP are peer-reviewed.

**International Journal of Livestock Production (IJLP)** (ISSN 2141-2448) is monthly (one volume per year) by Academic Journals.

**Contact Us** 

Editorial Office:	ijlp@academicjournals.org
Help Desk:	helpdesk@academicjournals.org
Website:	http://www.academicjournals.org/journal/IJLP
Submit manuscript online	http://ms.academiciournals.me/

### **Editors**

Prof. Carlos A. Gomez Nutrition Department, Faculty of Zootechnical -Universidad Nacional Agraria La Molina Peru

#### Dr. K.N. Mohanta

Fish Nutrition and Physiology Division, Central Institute of Freshwater Aquaculture, Indian Council of Agricultural Research (Ministry of Agriculture, Government of India), Kausalyganga, Bhubaneswar, 751 002, India.

#### Prof. Shaukat Ali Abdulrazak

National Council For Science and Technology P.O. Box 30623-00100, Nairobi, Kenya.

#### Dr. S.P. Muthukumar

Animal House Facility (B&N), Central Food Technological Research Institute, CSIR, Mysore - 570020, Karnataka, India.

#### Dr. Frederick Yeboah Obese Ruminant

Nutrition and Physiology, Department of Animal Science, College of Agriculture and Consumer Sciences, University of Ghana, Legon, Ghana.

#### Dr. Nicola Koper

Natural Resources Institute, University of Manitoba, Winnipeg, MB, R3T 2N2, (204) 474-8768, Canada.

#### Dr. Ramesh Khanal

Arkansas Children's Nutrition Center (ACNC), 1212 Marshall Street, Little Rock, AR 72205 USA. **Prof. Maher H. Khalil** College of Agriculture and Veterinary Medicine, Qassim University, Saudi Arabia .

**Dr. Ming-Che Wu** Taiwan Livestock Research Institute Taiwan.

#### **Dr. Ola Safiriyu Idowu** Department of Animal Science, Obafemi Awolowo University, 220005, Ile-Ife,

Nigeria.

Osun State,

#### Dr. Olubayo Reardon

Livestock sector, Ministry of Livestock Development, FAO (Sierra Leon) and FARM-Africa Kenya.

#### Dr. Sandip Banerjee

Department of Animal and Range Sciences, Hawassa University, Ethiopia.

#### Prof. Tchouamo Isaac Roger

Faculty of Agriculture, Department of Extension Education and Rural Sociology, University of Dschang, Dschang Cameroon.

#### Prof. Dale R. ZoBell

Department of Animal, Dairy and Veterinary Sciences, Utah State University, Logan, UT USA.

## **Editorial Board**

#### Dr. SHOOR VIR SINGH

Microbiology Laboratory, Central Institute for Research on Goats, Makhdoom, PO - FARAH, Dist. Mathura, UP, INDIA.

#### Dr. OSCAR IRAM ZAVALA LEAL

Centro Interdisciplinario de Ciencia Marinas Unidad Piloto de Maricultivos La Paz, BCS. Mexico

#### Dr. Ruheena Javed

Kurukshetra University Kurukshetra,Haryana, India.

#### Dr. Juarez Lopes Donzele,

Ph.D., Professor Department of Animal Science Universidade Federal de Viçosa (Federal University of Viçosa, Brazil). Brazil.

#### Dr. Daniella Jorge de Moura,

Ph.D., Assistant Professor School of Agricultural Engineering Universidade Estadual de Campinas (State University of Campinas, Brazil) Brazil. **Dr. Rita Flávia Miranda de Oliveira,** *Ph.D., Assistant Professor Department of Animal Science Universidade Federal de Viçosa (Federal University of Viçosa, Brazil), Brazil* 

#### Dr. Richard S. Gates,

Ph.D., Professor Agricultural and Biological Engineering Department University of Illinois at Urbana/Champaign, Urbana/Champaign, IL, USA

#### Dr. Angela R. Green,

Ph.D., Assistant Professor Agricultural and Biological Engineering Department University of Illinois at Urbana/Champaign, Urbana/Champaign, IL, USA.

#### Dr.Tugay AYAŞAN

East Mediterranean Agricultural Research Institute, Karatas Road, 01321, Yuregir/Adana Turkey.

## **International Journal of Livestock Production**

Table of Contents:Volume8Number1January2017

### **ARTICLES**

Evaluation of oestrus synchronization and mass artificial insemination service of dairy cattle in Mizan Aman area, Bench Maji zone, South West Ethiopia Tegegn Fantahun and Zelalem Admasu

Handling practices and microbial contamination sources of raw milk in rural and peri urban small holder farms in Nakuru County, Kenya Orwa J. D., Matofari J. W. and Muliro P. S.

5

1

## academic Journals

Vol. 8(1), pp. 1-4, January 2017 DOI: 10.5897/JJLP2016.0338 Article Number: BAB187562441 ISSN 2141-2448 Copyright ©2017 Author(s) retain the copyright of this article http://www.academicjournals.org/JJLP

International Journal of Livestock Production

Full Length Research Paper

## Evaluation of oestrus synchronization and mass artificial insemination service of dairy cattle in Mizan Aman area, Bench Maji zone, South West Ethiopia

#### Tegegn Fantahun\* and Zelalem Admasu

Department of Animal Science, Mizan-Tepi University, P. O. Box 260, Mizan-Teferi, Ethiopia.

Received 7 September, 2016; Accepted 22 December, 2016

The objectives of this study were to assess the hormonal response, conception rate, calving rate and perception of farmers towards the technology. From 220 cows and heifers brought by farmers for the services, 65% (143) that fulfilled the selection criteria were selected and injected with 2 ml of Cloprostenol. Data on the history of each heifers and cows, number of heifers and cows responsive to hormonal treatment, conception and calving rates were collected. The collected data were analyzed using descriptive statistics. The results of the finding showed that 91(63.64%) of cows and heifers were responsive to hormonal treatment. Majority 81(89.01%) of responding cows and heifers were inseminated. Finally, 11(13.58%) of calves were delivered. Oestrus response rate was relatively high, but conception rates and calving rates were very low. The lower percentages of conception rate which was observed in this study were associated with heat detection problems of farmers, distance from artificial insemination (Al) service centers, timing of insemination and poor husbandry practice of heifers and cows. To improve the effectiveness of the technology, there is a great need of skilled and experienced technician, and capacity building of farmers in heat detection and husbandry practices. Improvements in facilities and management should be necessary before implementing effective estrous synchronization and mass artificial insemination program.

Key words: Cloprostenol, cow/heifer, artificial insemination, oestrus synchronization.

#### INTRODUCTION

Livestock systems in developing countries are characterized by rapid change (Delgado et al., 1999; Thornton et al., 2007) and currently contributes about 30% of agricultural gross domestic product, with a projected increase to about 40% by 2030 (FAO, 2010).

The Ethiopian cattle population is estimated to be about 53.4 million, of which 55.2% are females. Out of total cattle population, 99.26, 0.64 and 0.1 percent are local, hybrid and exotic breeds, respectively (CSA, 2011). With an average lactation length of 6 months and an average

\*Corresponding author. E-mail: tegefanta@yahoo.com.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> daily milk production of 1.85 liters per cow, the total milk produced during the year 2010/11 was recorded to be 4.06 billion liters (CSA, 2011). Due to productivity of indigenous cattle, the country is still importing a significant amount of dairy products (Zijlstra et al., 2015).

Cattle breeding are mostly uncontrolled in Ethiopia making genetic improvement difficult (Azage et al., 1995). The total number of exotic and hybrid female cattle produced through the crossbreeding programme for decades in the country is quite insignificant indicating unsuccessful crossbreeding through artificial insemination (AI) (CSA, 2011; Desalegn, 2008; Sinishaw, 2004). Low pregnancy rate following artificial insemination in most African countries is attributed to poor semen quality, poor semen handling procedure, inadequate insemination skill, poor estrus detection and wrong time of insemination (Azage et al., 1995). The use of artificial insemination in Ethiopia is growing but oestrus detection is difficult owing to poorly expressed estrus of Zebu breeds (Mugerwa and Azage, 1991). To improve efficiency of artificial insemination practice in Ethiopia, hormonal synchronization of oestrus have been available for the past few years and have enjoyed success as a tool to make artificial insemination more practical(Azage et al., 2012; Gizaw et al., 2016). However, farmers expressed low satisfaction with the service, although evaluation of the technology by farmers is confounded with low conception rates (Gizaw et al., 2016).

In Ethiopia, attempts to improve the productivity of cattle have been made especially in the area of crossbreeding for the last decades but with little success (Aynalem, 2006). Hormonal oestrus synchronization could be used for increasing the probability of oestrus detection, much calving with feed availability and market demand for dairy products and increase pregnancy rates of dairy cattle (Azage et al., 2012; Lucy et al., 2004). There are different types of protocols available for synchronizing oestrus in cattle (Gizaw et al., 2016). In the study area, single injection of Cloprostenol followed by heat detection and artificial insemination protocol was used. It is important to evaluate the success and failure of the hormonal oestrus synchronization and mass artificial insemination programme so as to provide appropriate solutions in the future. Therefore, this research was conducted having as objectives to identify the hormonal response rate, conception rate and calving rate of cows and heifers and to assess the perception of farmers related with the service.

#### MATERIALS AND METHODS

#### Description of the study area

The study was conducted in Mizan Aman area which is situated in Bench Maji Zone south western part of Ethiopia. It is located at 585 km south west of Addis Ababa the capital of Ethiopia. Regarding the agro-Ecology of the zone, out of the total land size 28.042% is low land, 15.44% midland and 56.74% highland. The annual mean temperature ranges between 15.1 and 27°C and the annual mean rain fall ranges from 400 to 2000 mm (BMZFED, 2012).

#### Selection of experimental animals

From the Mizan Aman area, three sites (Addis Ketema, Kometa and Aman) were selected based on proximity to animal handling crush and cattle population. Out of 220 cows and heifers brought to the three sites, only 143(65%) (55 from Addis Ketema, 66 from Kometa and 22 from Aman) were selected. Among selected cows and heifers, 137(95.8%) were Zebu, 4(2.8%) were Sheko and 2(1.4%) were Cross breed. The average body weight of cows/heifer was 208.4 kg (range from 180 to 308). The average age of cows and heifer was 6.36 years range from 4 to 9 years.

The females which were diagnosed to be cycling with presence of a functional CL was determined through rectal palpation by AI technician were injected (2 ml) PGF2 $\alpha$  (Synchromate, Bremer Pharma GMBH, Germany, 1 ml solution of Synchromate contains cloprostenol 0.263 mg equal to cloprostenol 0.250 mg) intramuscular. The protocol used for the experiment was one single injection, heat detection and artificial insemination.

#### Data collection

The study was conducted from August 2014 to July 2015. Data on age of the cow and heifers, breed, body weight, date and time of hormone treatment, date and time of oestrus detection, date and time of artificial insemination, conception rate (pregnancy diagnosis was carried out at three months of post artificial insemination by rectal palpation) and delivery rate were recorded. Group discussion was also conducted at each site to assess the perception of farmers tow ards the technology.

#### Data analysis

The data were interred in Microsoft Excel, checked and analyzed by descriptive statistics using SPSS computer software program (version 17). Oestrus rate (Number of cow show ed oestrus/Number of cow s treated multiplied by 100) and conception rate (No. of cow s/heifers pregnant / No. of cow s/heifers inseminated multiplied by 100) were also calculated.

#### RESULTS

#### Hormonal response and insemination rate

The result of the finding showed that 91(63.64%) of cows and heifers were responsive to hormonal treatment. Majority 81(89.01%) of responding cows and heifers were inseminated, 2.19% of cows were aborted due to the drug effect as animal were at early stage of pregnancy and the remaining cows/heifer which did not show heat signs were not inseminated as shown in Table 1.

#### Conception and calving rate

The conception rate of 24.69% (20) was obtained, after three months pregnancy diagnosis. Finally, the calving rate was 13.58% (11). A total of 8 females and 3 males were delivered finally as presented in Table 1.

No.	Description	Adis Ketema		Kometa		Aman	
		Freq	%	Freq	%	Freq	%
1	Cows/heifer synchronized	55		66		22	
2	Cows/heifer responsive	36	65.45	41	62.12	14	63.63
3	Cows/heifer inseminated	36	100	41	100	4	28.6
4	Calf delivered	4	8.3	6	14.63	1	7.14
5	Cows/heifer sold	2	3.63	5	7.57	0	0
6	Cows/heifer slaughtered	5	9.09	1	1.51	0	0
7	Cows aborted	0	0	1	2.77	0	0
8	Cows/heifer dead	1	33.33	0	0	0	0

Table 1. Oestrus synchronization response, artificial insemination and pregnancy rates of cows and heifers in the study area.

#### Perceptions of the farmers towards the technology

After group discussion with farmers in each site, they had interest to get the services to have improved breed. Cows and heifers were travelled more than 5 km to get the service. In the study area, only few AI technicians were serving large population of cattle and there were no effective regular AI service. Farmers, in the area, were not aware of hormonal oestrus synchronization protocols and AI technology, which contributed in the poor efficiency of the services. Lack of awareness of associated with some farmers during group discussion were immediately mix cows and heifers with other herds after hormonal injection, long distance trucking of cows and heifers, cows and heifers were not brought at the right time for insemination and poor management practices. In general, farmers' perceptions with hormonal oestrus synchronization technology were variable and the satisfaction of them determined by calving rates. Therefore, those farmers that got calf develop positive perception towards the technology and satisfied than others.

#### DISCUSSION

As compared with the current finding, using single injection of prostaglandin F2α (Lutalyse) protocol different response rate was reported in different part of the country, higher oestrus responses rate were reported by Azage et al. (2012) who reported 97.7% in Hawassa-Dale milk shade and 100% in Adigrat-Mekelle milk shade areas. Adebabay et al. (2013) reported an oestrus rate of 89.3% in Bahir Dar milkshed; 72.3 and 92.17% oestrus rate reported in West Shoa zone by Bainesagn (2015) and Girmay et al. (2015) in Wukro Kilte Awulaelo district, in Northern Ethiopia, respectively. Moreover, using the same protocol with the current study, 84.2% oestrus rate was reported in eastern zone, of Tigray region, Ethiopia (Tadesse, 2015).

The conception rate obtained in this study was higher than 13.7% reported by Adebabay et al. (2013) in Bahir

Dar milk shed area. In contrast to this finding, in Hawassa-Dale milk shade, 57.7% and in Adigrat- Mekelle milk shade 61.7% of pregnant animal was reported by Azage et al. (2012); 32.17% pregnancy rate was reported in Wukro Kilte Awulaelo district (Girmay et al., 2015); 59.6% conception rate was reported in eastern zone, of region, Ethiopia (Tadesse, 2015). Factors Tigray associated with this lower rate of pregnancy might be related with timing of artificial insemination, feeding efficiency management. of heat detection, early embryonic mortality and presence of ovarian cyst which are all known to negatively affect fertility. Factors affecting embryonic/fetal loss are numerous and include genetic abnormalities, fescue toxicosis, plant toxins, excess protein, heat stress, reproductive diseases, an effect of the sire, and handling or transportation stress (Smith et al., 2011)

#### Conclusions

As revealed by results of this study, using single injection prostaglandin/Cloprostenol/ was effective to synchronize cows and heifers. Cows and heifers come to heat within short period of time which reduces calving interval. Oestrous response rate was relatively high, but conception rates and delivery rate were very low. To improve effectiveness of the technology, skilled and experienced technicians as well as capacity building of farmers in heat detection and husbandry practices are of major concerns.

#### **Conflicts of Interests**

The authors have not declared any conflict of interests.

#### ACKNOWLEDGEMENT

We would like to provide our heartfelt gratitude to farmers who participated in the study. Finally, they would like to extend their thanks to Mizan Tepi University for financial support to conduct the research project.

#### **REFERENC ES**

- Adebabay K, Getinet Z, Yeshwas F,Temesegen A, Azage T (2013). Prostaglandin (PGF2 α) based oestrous synchronization in postpartum local cows and heifers in Bahir Dar milkshed. Int. J. Pharm. Med. Bio. Sci. 2(4):37-43.
- Aynalem H (2006). Genetic and economic analysis of Ethiopian Boran cattle and their crosses with Holstein Friesian in central Ethiopia. PhD thesis. National Dairy Research Institute (I.C.A.R) karnal-132001 (Haryana), India. P 197.
- Azage T, Awet E, Asrat T, Dirk H (2012). Technological options and approaches to improve smallholder access to desirable animal genetic material for dairy development: IPMS Experience with hormonal oestrus synchronization and mass insemination in Ethiopia. Trop. pp. 19-21.
- Azage T, Kassa T, Mugerw a E M (1995). Aspects of bull production with emphasis on cattle in Ethiopia. Sperm production capacity and semen characteristics. In: Proceeding of the Third National Conference of Ethiopian Society of Animal production, pp. 83-99.
- Bainesagn W (2015). Assessment of breeding practices and evaluation of estrus synchronization and mass insemination technique in dairy cattle in West Shoa zone. Msc thesis, Haramaya University, Ethiopia.
- Bench Maji Zone Finance and Economic Development (BMZFED) (2012). Annual report of Bench Maji zone Finance and Economic Development Main Department. Mizan-Teferi, Ethiopia.
- Central Statistical Authority (2011). Agricultural sample survey 2010/11. Report on livestock and livestock characteristics. Central Statistical Authority (CSA), Addis Ababa, Ethiopia 2(502).
- Delgado C, Rosegrant M, Steinfeld H, Ehui S, Courbois C (1999). Livestock to 2020: the next food revolution. Food, Agriculture and the Environment. IFPRI/FAO/ILRI, Washington, DC, USA. Discussion Paper. P 28.
- Desalegn GM (2008). Assessment of problems/constraints associated with artificial insemination service in Ethiopia. MSc thesis, Addis Ababa University, Ethiopia.
- Food and Agriculture Organization (2010). Breeding strategies for sustainable management of animal genetic resources. Animal Production and Health Guidelines, Food and Agriculture Organization (FAO) of the United Nations, Rome, Italy.
- Girmay G, Berihu G, Bahlibi W (2015). The effect of one shot Prostaglandin on Estrus Synchronization of Local and Holstein Friesian Cattle in and around Wukro Kilte Aw ulaelo District, Northern Ethiopia. J. Bio. Agric. Health 5(7):99-105.

- Gizaw S, Tesfaye Y, Mekuriaw Z, Tadesse M, Hoekstra D, Gebremedhin B, Tegegne A (2016). Oestrus synchronization for accelerated delivery of improved dairy genetics in Ethiopia: Results from action research and development interventions. International Livestock Research Institute (ILRI). Nairobi, Kenya. LIVES Working Paper 12.
- Lucy MC, McDougall S, Nation DP (2004). The use of hormonal treatments to improve the reproductive performance of lactating dairy cows in feedlot or pasture-based management systems. Anim. Reprod. Sci. 82(83):495-512.
- Mugerwa EM, Azage T (1991). Reproductive performance in Ethiopia zebu (Bos indicus) cattle: Constraint and impact on production. pp. 16-28 In: Proceeding on the 4th National Livestock Improvement Conference. Ethiopian Institute of Agricultural Research, Addis Ababa. Ethiopia.
- Sinishaw W (2004). Study on semen quality and field efficiency of AI bulls kept at the National Artificial Insemination Center. MSc thesis, Addis Ababa University, Faculty of Veterinary Medicine, Debre Zeit.
- Smith MF, Perry GA, Atkins JA, Jinks EM, Pohler KG, Patterson DJ (2011). Keys to a successful estrus synchronization and artificial insemination program. Proceedings, Applied Reproductive Strategies in Beef Cattle August 31 – September 1, Joplin, MO.
- Tadesse G (2015). Effects of prostaglandin administration frequency, artificial insemination timing and breed on fertility of dairy cows and heifers in eastern zone, of Tigray region, Ethiopia. MSc thesis, Mekelle University, Ethiopia.
- Thornton P, Herrero M, Freeman A, Mw ai O, Rege E, Jones P, McDermott J (2007). Vulnerability, climate change and livestock – research opportunities and challenges for poverty alleviation. International Livestock Research Institute (ILRI), Nairobi, Kenya. SAT eJournal| ejournal. icrisat. org 4(1).
- Zijlstra J, Berhanu T, Vernooij A, Boere A, Jvd L (2015). Business Opportunities Report Dairy #2 in the series written for the Ethiopian Netherlands business event. Rijswijk, The Netherlands.

## academic Journals

Vol. 8(1), pp. 5-11, January 2017 DOI: 10.5897/JJLP2016.0318 Article Number: 62B12062457 ISSN 2141-2448 Copyright ©2017 Author(s) retain the copyright of this article http://www.academicjournals.org/JJLP

International Journal of Livestock Production

Full Length Research Paper

## Handling practices and microbial contamination sources of raw milk in rural and peri urban small holder farms in Nakuru County, Kenya

Orwa J. D.<sup>1</sup>, Matofari J. W.<sup>1</sup>\* and Muliro P. S.<sup>1</sup>

Department of Dairy and, Food Science and Technology Faculty of Agriculture, Egerton University, Nairobi, Kenya.

#### Received 23 June, 2016; Accepted 8 September, 2016

The cow, the milking and milk handling procedures at the farm level expose the milk to potential risk of contamination with spoilage microorganisms. Milk contamination if not prevented will lead to milk losses along the dairy value chain. The objective of this study was therefore to identify the risk factors associated with contamination of milk with spoilage microorganisms at the farms in rural and peri urban in Nakuru County Kenya. A survey was conducted using a pre-tested semi structured questionnaires (250) and an observation checklist to identify the risk factors. A total of 560 samples obtained from the following identified contamination sources; the udder, hands, milking and bulking containers and water sources were analyzed for total viable counts (TVC), Coliform counts (CC), thermophillic bacteria counts (ThBC) and psychrophilic bacteria counts (PBC). The results from the survey showed that only 11% of rural farmers practiced hand and udder drying compared to 50% in peri-urban. Water treatment by either chlorination or boiling was done by 11% in rural and 30% in periurban respectively. Regression of risk factors versus farm gate milk from viable colony counts, showed that udder swabs were the highest source of contamination of milk (r =2.73). In the rural, hands of milking personnel recorded the highest for TVC (log<sub>10</sub> 3.7 CFU/ml). It is evident from the results that effective udder cleaning and observation of high personal hygiene by the hand milkers may reduce the risk of microbial contamination in both systems of milk production.

Key words: Risks, handling practices, contamination, rural, peri urban.

#### INTRODUCTION

Livestock contributes about 50% to the agricultural Gross Domestic Product (GDP) in Kenya with dairy production contributing up to 33% of this (Lore et al., 2005). Milk production in Kenya is mainly from cattle, camel and goats. Dairy cattle however account for over 70% of national milk. The main dairy breeds include Friesian, Ayrshire, Jersey, Guernsey their crosses and indigenous cows. Smallholder dairy farmers dominate the dairy industry by accounting for over 75% at the production level (FAO, 2011).

Contamination of milk however begins at the farm during and after harvest (Kornacki and Johnson, 2001).

<sup>\*</sup>Corresponding author. E-mail: jmatofari@gmail.com.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u>

Research has showed that most post- harvest milk losses are experienced in small scale dairy farms and at the farm level (Muriuki, 2003; Lore et al., 2005; FAO, 2011). Farm losses have been recorded to be highest in several countries. Kenya recorded the highest volume in losses at the farm in east Africa, standing at 54.2 million liters compared to 8.4 million liters, 28.6 million liters, 46.4 million liters in Uganda, Ethiopia and Tanzania respectively (Lore et al., 2005)

On farm dairy losses occur in three main forms; spillage spoilage and forced consumption. Spillage is caused by poor roads during transportation; spoilage is caused by spoilage microorganisms which produce lactic acid increasing the milk acidity above the accepted levels. When the milk fails the alcohol test it is rejected there after it is returned to the farm. At the farm the milk is forcefully consumed, thrown away, or sold at throw away prices leading to economic loss (Muriuki, 2003; FAO. 2011). Losses at the farm has been reported by World Bank to cost the farmers 2\$ each month in developing countries (Bonfoh et al., 2003; Paola et al., 2013). Losses have also been attributed to by lack of adequate animal health control, inadequate training among farmers and farm employees on milk hygiene (Chye et al., 2004; Chizari et al., 2008; Paola et al., 2013).

Harvesting of milk which basically takes place at the farm faces many sources of contamination. The animal itself is a risk factor. If the cow is not healthy, then the milk is likely to be contaminated with microorganisms such as Staphylococcus, Streptococcus, enteric bacteria among others in cases of subclinical mastitis at the udder. Other commensal and pathogenic microorganisms have also been isolated from the udder. Traditional pre milking and post milking procedures used during harvesting are risk factors more so when milking is done in open fields in non-controlled environments. The milking environment in small scale farms are sometimes characterized by dust and faeces. The milking hands of personnel, milking containers and bulking containers are contact surfaces of the milk posing as risks for milk contamination. The water used at the farm during cleaning of the udder, hands and equipment has been considered a factor in milk contamination (Ingawa et al., 1992; Teka, 1997; Walstra et al., 1999; Kornacki and Johnson, 2001; Petrovick et al., 2006; Visser et al., 2007; Coorevitis et al., 2008; Kumar et al., 2012; Al-Hubeatyet al., 2013; Gleeson et al., 2013; Matofari et al., 2013). The major milk spoilage bacteria that have been isolated from raw milk include; coliforms, lactic acid bacteria (LABs), psychrotrophic bacteria (Pseudomonas spp.) and thermophillic bacteria (Bacillus spp.) (Griffiths and Phillips, 1990; Bareeda, 2012; Gleeson et al., 2013; Paola et al., 2013; Mesfine, 2015).

There is a deficiency in information on the microbiological quality of these risk factors pointing out the most responsible source of contamination of milk

after harvest at the farm. The aim of this study was to assess the risk factors in small scale farms associated with contamination of milk with spoilage microorganisms while at the same time profile the microbiological quality of these risk factors. The outcome of the study is expected to build on previous studies and be a useful source in developing mitigation measures to curb losses due to spoilage at the farm.

#### MATERIALS AND METHODS

#### Study site

The study was carried out in Nakuru county Kenya where dairy farming is thriving. Nakuru county is a Kenyan highland found in rift valley where dairy milk production is highest in the country (Muriuki, 2003). Two locations were selected to capture rural and peri urban farm characteristics. Olenguruone sub-county is a rural setting which lies about 35°40' 60"E and 0°34' 60"S while Bahati-Wanyororo Sub-County is a peri-urban setting next to Nakuru town which lies about 36° 40'60"E and 0°40' 60"N. Small scale dairy farmers were targeted because thy account for over 80% of milk producers in the country (FAO, 2011).

#### Conduct of survey

A cross sectional survey was done using a pretested questionnaires and observation checklist. In the rural setting 150 questionnaires were administered and 100 in the peri-urban. The questionnaire targeted farm characteristics highly associated with milk contamination with spoilage microorganisms. Some of the characteristics included; method of grazing, water sources, method of water treatment, waste disposal, training of farm personnel, method of cleaning milking and bulking containers, types of material for the containers. The observation checklist was used at milking time to assess the hygiene practices followed. Pre milking practices such as; udder and hand washing, pre-dipping, hand and udder wiping, type of material used for drying hands and udder. Post milking practices sought were; post dipping, and udder drying. Stratified random sampling was done in selection of farms for the survey since not all farmers had dairy cows and out of which only those willing to participate in the practice were selected.

#### Sample collection for microbiological analysis

Contamination sources identified from the survey (hands, udders, water source, milking and bulking containers) were sampled for microbiological analysis. Samples were collected early in the morning during milking time. Sample collection begun after the udder was cleaned and ready for milking and the milking personnel had done the necessary pre milking preparation. Sterile cotton swabs wrapped in splint wood sticks were used in swabbing hands and udder. A surface area of 38.5 cm<sup>2</sup> from both hands of the milking personnel was swabbed. The swab was then immediately transferred into a sterile Stuart Transport Medium (Oxoid) in a screw cap Bijou bottle. The handle stick was broken while the swab remained in the transport medium. The cap of the bottle was then put back and transferred to the cool box. The teat of the udder was swabbed from the attachment of the teat to the udder downwards while avoiding contact with hair on the udder (Kumar, 2012). The four teats per cow were swabbed and transferred to the same bijou bottle.

Milk (50 ml) from all the quarters of the udder was collected in a



Farm Practices and charcateristics

Figure 1. Graph of farm practices and characteristics of farms in the rural and peri urban locations.

50 ml sterile sampling bottle. Milking containers and bulking containers at the farm gate were rinsed with 100 ml sterile water and the volumes of the containers were recorded. Milk at the farm gate in the bulking container was also sampled in sterile sampling bottles. Approximately 400 ml of water source used at the farm was sampled in a 500 ml sterile sampling bottle. The swabs, milk and water samples were transported in a cool box with ice bags at 8 to 10°C to the laboratory in six hours. In the peri urban, 30 farms were visited for sample collection which provided a total of 210 samples while in the rural, 50 farms were visited and this provided 350 samples making a total of 560.

#### **Microbiological analysis**

Examination of samples for total viable counts (TVC), coliform counts (CC), Thermophillic bacterial counts (ThBC) and psychrophilic bacterial counts (PBC) were done by standard procedures of International Dairy Federation (IDF), East African Standards of milk examination (EAC, 2006) and ISO (International Standard of Organisation). TVC was incubated at 32°C for 48 h (EAS 67:2000 (4.2.1) EAC 2006) in Plate count agar (OXOID). Colifrom counts were incubated at 30°C for 48 h (ISO, 2006) in MacConkey agar (OXOID). Psychrophillic bacteria was incubated at 6.5°C for 10 days (IDF) using Plate count Agar while thermophillic bacteria were incubated at 55°C for 48 h (Abdul-Hadi et al., 2014).

#### Data analysis

Data collected by survey questionnaires was used to determine risk factors. Data was analyzed using SPSS (Statistical Package for social scientists) version 20. A cross tabulation was done between the risk factors and location. Values obtained from the 100 ml rinse from milking and bulking containers was divided by the volume of the corresponding container to determine contamination (colony forming units per ml, CFU ml<sup>-1</sup>). The container volumes were not the same; however the bacterial counts were not corrected for this variation (Bonfoh et al., 2003). Microbial count data was first transformed to logarithmic values (log  $_{10}$ ) before subjection to statistical analysis.

The general linear model of SAS version 9.1.3 (SAS proc glm) was used to analyze milk microbial quality and the microbial quality of contamination sources. Mean comparison was done by the Fisher's least significant difference (LSD) when analysis of variance showed significant difference in means. Statistical difference was determined at 95% confidence level. The microbiological quality of milk was regressed versus the risk factors determined (udder swabs, hand swabs, milking container, bulking container and water source) to identify the risk factor which contributes highest to farm gate milk quality.

#### RESULTS

#### **Risk factors**

From the survey questionnaire and observation checklist, none of the farms visited in both rural and peri urban practiced machine milking. Hand washing was practiced by all farmers in peri urban. Drying of hands and udder was practiced by 11% of farmers in rural and 50% in peri urban. Plastic milking containers were 60% in peri urban and 84% in rural location (Figure 1). Cross tabulation of risk factors practices between location showed that lack of hand drying was significantly different (P=0.007). Plastic milking and bulking containers were significantly different (p=0.04 and p=0.03 respectively) between locations. Lack of water treatment was practiced by 60% in rural and 80% in peri urban this was significantly different (p=0.008).

#### Microbiological quality of contamination sources

From the survey the risk sources of contamination to milk contamination with spoilage microorganisms included udder, hands, milking containers, bulking containers and

Missel		Risk factors						
count	Location	Milk directly from udder	US	HS	MCR	BCR	ws	Milk at the farm gate
TVC	Rural	5.2±0.3 <sup>a</sup>	3.4 ±0.2 <sup>a</sup>	3.0±0.3 <sup>a</sup>	3.3±0.2 <sup>a</sup>	2.4±0.5 <sup>a</sup>	2.6±0.2 <sup>a</sup>	5.7±0.5 <sup>ª</sup>
	Peri urban	4.8±0.5 <sup>a</sup>	3.9±0.2 <sup>a</sup>	3.7±0.2 <sup>b</sup>	4.4±0.3 <sup>b</sup>	2.1±0.1 <sup>a</sup>	2.5±0.6 <sup>a</sup>	5.2±0.4 <sup>ª</sup>
СС	Rural	3.2±0.8 <sup>a</sup>	2.7±0.3 <sup>a</sup>	3.3±0.4 <sup>a</sup>	1.3±0.4 <sup>a</sup>	1.6±0.7 <sup>a</sup>	1.5±0.3 <sup>a</sup>	3.9±0.8 <sup>a</sup>
	Peri urban	4.2±0.4 <sup>b</sup>	3.5±0.3 <sup>b</sup>	3.6±0.2 <sup>b</sup>	1.1±0.5 <sup>a</sup>	1.1±0.2 <sup>b</sup>	1.8±0.4 <sup>a</sup>	4.7±0.3 <sup>b</sup>
ThBC	Rural	5.0±0.4 <sup>a</sup>	2.8±0.3 <sup>a</sup>	2.8±0.3 <sup>a</sup>	2.5±0.3 <sup>a</sup>	2.1±0.6 <sup>a</sup>	2.2±0.3 <sup>a</sup>	$5.2\pm0.5^{a}$
	Peri urban	2.7±0.5 <sup>b</sup>	3.2±0.3 <sup>a</sup>	3.4±0.2 <sup>b</sup>	1.1±0.3 <sup>b</sup>	2.1±0.4 <sup>a</sup>	1.4±0.3 <sup>b</sup>	2.8±0.6 <sup>b</sup>
PBC	Rural	$5.5\pm0.3^{a}$	3.3±0.3 <sup>a</sup>	1.8±0.5 <sup>ª</sup>	3.3±0.2 <sup>a</sup>	1.9±0.9 <sup>a</sup>	2.6±0.4 <sup>a</sup>	6.1±0.3 <sup>a</sup>
	Peri urban	3.7±0.4 <sup>b</sup>	3.2±0.3 <sup>a</sup>	3.7±0.2 <sup>b</sup>	2.5±0.4 <sup>b</sup>	2.4±0.4 <sup>b</sup>	2.9±0.2 <sup>a</sup>	4.7±0.8 <sup>b</sup>

**Table 1.** Table of microbial counts (Means  $\pm$  SE) of risk factors and milk drawn directly from the udder and at the farm gate.

US, udder swabs; HS, hand swabs; MCR, Milking container rinse; BCR, bulking container rinse; WS, water source. TVC, total viable counts; CC, coliform counts; ThBC, thermophilic bacterial counts; PBC, psychrophillic bacterial count. Means followed by the same letter in a column within a row are not significantly different at (P > 0.05).



**Figure 2.** Comparison of bacterial counts in raw milk drawn directly from the udder and milk at the farm gate in the rural. TVC, Total viable counts; CC, coliform counts; ThBC, thermophillic bacterial counts; PBC, psychrotrophic bacterial counts.

water sources. Udder swabs recorded the highest counts in TVC (log <sub>10</sub> 3.4 CFU/ml) in rural while milking container recorded the highest (log<sub>10</sub> 4.4 CFU /ml) in peri urban. Hands and udder recorded the highest counts in coliform for both locations. Water source recorded the highest in PBC in peri urban location while milking container rinses recorded a significantly (p≤0.05) lower value for ThBC in peri urban. There was a steady rise in microbial counts between the udder to the farm gate in all microbial counts evaluated (Table 1). Increase in TVC from the udder to the farm gate was 0.5 log cycle in rural (Figure 2). A significant increase in coliform count was recorded between the udder and farm gate in rural and peri urban milk (Figure 3). Regression coefficients were derived from the formula below to determine the most responsible sources of different microbial types in milk at the farm gate. Where Y represented each microbial type in farm gate milk (TVC/CC/ThBC/PBC) and was regressed against the microbial type of each risk factor (US, HS, MCR, BCR and WS) evaluated and  $X_1$  to  $X_5$  are the regression coefficients of the respective risk factors.

$$Y = \beta o + X_1 US + X_2 HS + X_3 MCR + X_4 BCR + X_5 WS + E_i$$

Bulking containers showed the highest regression coefficient value in peri urban TVC of farm gate milk. Hands, udder and water source were the highest



**Figure 3.** Comparison of bacterial counts in raw milk drawn directly from the udder and milk at the farm gate in the peri urban. TVC, Total viable counts; CC, coliform counts; ThBC, thermophillic bacterial counts; PBC, psychrotrophic bacterial counts.

Table 2. Regression coefficients of risk factors versus farm gate milk.

Microbial type	Lesster	Risk factors						
	Location	US	HS	MCR	BCR	WS	Constant	
TVC	Rural	2.73*	2.63*	0.18	0.05	0.12	4.84	
	Peri urban	0.87	1.15	1.19	1.51*	0.60	3.29	
CC	Rural	0.83	1.46*	0.74	0.16	0.88*	5.34	
	Peri Urban	0.58	0.36	0.04	0.31	0.16	4.46	
ThBC	Rural	0.22	0.93*	0.02	0.83	0.59	1.85	
	Peri urban	0.22	0.31	0.37	0.17	0.08	4.78	
PBC	Rural	0.14	0.24	0.48*	0.02	0.01	4.42	
	Peri urban	0.46	0.62*	0.55	0.36	0.35	4.39	

US, udder swabs; HS, hand swabs; MCR, milking container rinse; BCR, bulking container rinse; WS, water source. TVC, Total viable counts; CC, coliform counts, ThBC, Thermophilic bacterial counts; PBC, Psychrophillic bacterial count. \* Regression coefficient significant at (P < 0.05).

contributors to coliform counts in rural farm gate milk. In peri urban udders were the highest contributors to coliform counts (Table 2).

#### DISCUSSION

The study established that farm practices which predisposed milk to microbial contamination included; lack of hand and udder washing, or washing without drying. A similar trend was observed in the peri-urban where 50% practice hand and udder drying (Figure 1). Without drying of hands and udder after washing becomes a risk because the water used in washing the udder and hands will drip in the milking container, mixing with the milk. The excess water from hands and udder if not dried off caries microorganisms from hands and udder in unhygienic conditions contributing to high microbial count in milk (Hogan et al., 1979; Gulton et al., 1984; Islam et al., 2009). It was reported that milking in a dry environment provides a significant reduction in microbial load in milk. Thus just washing hands and udder is not as effective as following the procedure with drying of the surfaces with a material like a towel (Islam et al., 2009).

Udder swabs in peri urban recorded high counts in TVC compared to their rural counterparts (Table 1). Due to the small pieces of land in peri-urban compared to the rural

areas, most farmers opt to practice zero grazing. Zero grazed animals stay in one place the whole day and are likely to have dirty udders due to defaecation in the same spot they feed and spend the night. The proximity of the udder and the rectum of the cow cause easy cross contamination from faecal coliform and other bacteria (Islam et al., 2009). With these factors, compared to the rural where free range grazing was mostly practiced due to availability of land, the hygiene of the udder was better than in peri urban.

Water treatment by either boiling or chlorination was more common in the peri urban than in the rural location (Figure 1). The microbial load in milk from rural areas where minimal water treatment was done, reported high cumulative microbial counts (Figure 3) than peri urban water source. The microbiological quality of water used during milking, udder preparation, and equipment cleaning in the farm play an important part in microbial load of raw milk. This showed that water hygiene is an important aspect of microbiological quality of milk. This water easily contaminates the milk especially where udder and hands drying after washing is not practiced. Previous studies have reported the same findings (Ingawa et al., 1992; Visser et al., 2007; Matofari et al., 2013).

Plastic milking containers which predominated the rural farms contributed to high microbial counts in the rural areas than the peri urban farms. Rinses from milking containers in the rural recorded the highest in total viable counts (Table 1). Plastic containers have been proven to contain micro -pores which facilitate the formation of biofilms and are therefore difficult to clean and become sources of contamination especially for psychrotrophic and thermophillic bacteria (Bereda et al., 2012; Mesfine et al., 2015). Methods of containers cleaning also vary from use of hot water, scouring material and detergent types. Since no standards methods exist in cleaning these containers are not controlled and therefore remain risks to milk contamination (Wafula et al., 2016).

Bulking containers had a significant regression coefficient value in total viable counts in peri urban unlike milking containers in both locations (Table 2). Milking containers in both locations had a wide opening to reduce spillage during milking from the udder. This property also helps in reducing biofilms due to ease of cleaning. The cleaning material can easily reach all parts of the container effectively. However, the wide opening of the milking container poses a risk of contamination from the milking environment which is always contaminated with cow dung. Bulking containers are however placed away from the milking area and do not get contamination from this kind of environment. Bulking containers are characterized with small openings to reduce spillage during transportation; however this property makes them difficulty to clean since not all areas are easily reached by cleaning material. This characteristic is a risk factor since the containers become hard to clean and promote the development of biofilms (Kaindi et al., 2011).

The micro-flora of milk at the farm gate is as a result of the contamination it acquires the moment it leaves the udder. Milk drawn directly from the udder had lower readings of TVC compared to the farm gate translating to 8.3% a percentage increase of in the rural which is a 0.5 log cycle. Hygiene in the peri-urban area was generally high compared to rural hence the high increase in microbial load in milk between the udder and the farm gate. This is because hand washing, udder washing and drying of the same was mostly practiced in the peri-urban compared to rural. Water treatment by boiling and chlorination was also practiced more in peri-urban compared to the rural counterparts. Other studies have reported lower counts in milk where proper pre milking and post milking practices were carried out targeting hands and udder (Hogan et al., 1979; Gulton et al., 1984; Islam et al., 2009: Odongo et al., 2016)

From the regression, the highest source of contaminant was the personnel followed by udder and bulking containers (Table 2). Lack of hand drying, zero grazing and proximity of the udder to the rectum are reasons for the high correlation between hands, udder and the microbiological quality of milk Hands and udder hygiene are majorly affected by pre-milking procedures and water quality which have shown in this study as being substandard. Mitigation measures in reducing microbial load and improving farm hygiene should target the practices associated with personnel activities, pre and post milking practices udder washing and drying before milking and using boiled or treated water or detergents to wash hands udder and containers. The microbiological quality and safety of milk is determined by handling and hygiene practices of the farm and the milk. Hygiene milking and post milking practices will ensure a low microbial count in milk with a longer shelf life (Petrovick et al., 2006; Kornacki and Johnson, 2001; Walstra et al., 1999).

#### Conclusion

Failure to observe high hygiene during milking and milk handling will expose milk to potential risk of microbial contamination. Udder of the cow is the highest source of contributor to milk contamination immediately it leaves the animal. It is evident from the study that effective udder cleaning and observation of high personal hygiene of the milking hands may reduce the risk of microbial contamination in both systems of milk production. Total microbial load in raw milk at the farm is highly contributed to by hands of milking personnel, udder swabs and bulking containers. Coliform counts are contributed to majorly by water source at the farm and milker's hands. Thermophilic bacterial counts are highly contributed to by hands majorly while psychrophilic bacterial counts are significantly affected by milking containers. The study was limited to microbial groups and did not identify specific microorganisms in terms of species. In future studies this aspect is recommended.

Most important is the need to train farmers and farm employees on the importance of farm hygiene especially where hand milking is involved. Resources should be directed towards increasing the knowledge base of farmers on the significant influence of microbial contamination at the farm to overall hygiene within the rest of the value chain in terms of safety and shelf life. The farmers should be encouraged to carry out outlined hygienic practices which include effective cleaning of hands, udder followed by proper drying. Water used at the farm should undergo treatment before use for milking preparations. The farmer should milk in a clean environment free of cow dung. Avoiding of calf suckling and tying the cow's tail during milking are practices which would reduce contamination of milk.

Abbreviations. HS, hand swabs; US, udder swabs; MCR, milking container rinse; BCR, bulking container rinse; WS, water source.

#### **Conflict of Interests**

The authors have not declared any conflict of interests.

#### ACKNOWLEDGEMENTS

This work could not be possible without aid from the Federal Ministry of Education and Research (BMBF) in Germany through the RELOAD (Reducing Losses Adding Value) project. The authors would also like to thank the farmers and cooperative societies of Olenguruone and Wanyororo who collaborated to make the research successful.

#### REFERENCES

- Abdul-Hadi H, Nadia I, Aysar S (2014). Prevalence of Thermophiles and Mesophiles in Raw and UHT milk. Int. J. Anim. Vet. Adv. 6(1):23-27.
- Bereda A, Yilma Z, Nurfeta A (2012). Hygienic microbial quality of raw whole cow's milk produced in Ezhadistrict of the Gurage zone, Southern Ethiopia. Wudpecker J. Agric. Res. 1(11):459-465.
- Chizari M, Jannat S, Abbasi S (2008). Role of extension in developing dairy farmers knowledge toward milk quality in Golpayegan township, Iran. American- Eurasian J. Agric. Environ. Sci. 3: 333-338.
- Chye F, Abdullah A, Ayob MK (2004). Bacteriological quality and safety of raw milk in Malaysia. Food Microbiol. 21:535-541.
- East African Community (EAC) (2006). ICS 67-2000 East African Standards. Raw cow Milk Specification.
- Gleeson D, O'Connell A, Jordan K (2013). Review of potential sources and control of thermoduric bacteria in bulk-tank milk. Irish J. Agric. Food Res. 217-227.

- Griffiths MW, Phillips JD (1990). Incidence, source and some properties of psychrotrophic Bacillus spp found in raw and pasteurized milk. Int. J. Dairy Technol. 43(3):62-66.
- Hogan H, Galton WA, Jr. OM, Adkinson RE, Pankey JW (1979). Effect of Pre-milking Udder Preperation on Milk Quality. Dairy Sci. Abs. 41:869.
- Hutchison ML, Thomas DJI, Moore A, Jackson DR, Ohnstad I (2005). An evaluation of raw milk microorganisms as markers of on-farm hygiene practices related to milking. J. Food Prot. 68(4):764-772.
- Ingawa KH, Adkinson RW, Gough RH (1992). Evaluation of a Gel Teat Cleaning and Sanitizing Compound for Premilking Hygiene 1, 2. J. Dairy Sci. 75(5):1224-1232.
- Islam MA, Islam MN, Khan MAS, Rashid MH, Obaidullah SM (2009). Effect Of Different Hygenic Condition During Milking On Bacterial Count Of Cows' Milk. Bangladesh J. Anim. Sci. 38(1-2):108-114.
- ISO (2006). Microbiology of food and animal feeding stuffs. Horizontal method for the enumeration of coliforms. Colony-count technique. ISO 4832:2006. International Organization for Standardization, Geneva.
- Kaindi DWM, Schelling E, Wangoh J, Imungi JK, Farah Z, Meile L (2011). Microbiological Quality of Raw Camel Milk across the Kenyan Market Chain. Global Science Books. (1):79-83.
- Kalogridou-Vassiliadou D (1992). Biochemical activities of Bacillus species isolated from flat sour evaporated milk. J. Dairy Sci. 75(10):2681-2686.
- Kornacki JL, Johnson JL (2001). Enterobacteriaceae, coliforms, and Escherichia coli as quality and safety indicators. Compendium of methods for the microbiological examination of foods. 4:69-82.
- Kumar AV, Rao LV, Kumar MK, Srinu B, Rao TM (2012). Efficacy of udder disinfectants on reduction of bacterial load and certain pathogens of public health significance. J. Microbiol. Biotechnol. Res. 2(1):147-151.
- Matofari JW, Shalo PL, Younan M, Nanua JN, Adongo A, Qabale F, Misiko BN (2013). Analysis of microbial quality and safety of camel (Camelus dromedarius) milk chain and implications in Kenya. J. Agric. Ext. Rural Dev. 5(3):50-54.
- Mesfine S, Feyera T, Mohammed O (2015). Microbiological Quality of Raw Cow's Milk from Four Dairy Farms in Dire Dawa City, Eastern Ethiopia. World J. Dairy Food Sci. 10(1):09-14.
- Muriuki H (2003). Milk and Dairy Products, Post-harvest Losses and Food Safety in Sub-Saharan Africa and the Near East: A Review of the Small Scale Dairy Sector–Kenya. Rome, Italy: Food and Agricultural Organization.
- Odongo NO, Lamuka PO, Matofari JW, Abong GO (2016). Risk factors associated with the post-harvest loss of milk along camel milk value chain in isiolo county Kenya. Afr. J. Agric. Res. 11(8):674-682.
- Paola B, Anna FAC, Simone S, Sara B, Casimiro C (2013). Microbiological survey of milk and dairy products from a small scale dairy processing unit in Maroua (Cameroon). J. Food Control Elsevier. 32:366-370.
- Petrović M, Pavičić Ž, Tomašković A, Cergolj M (2006). Effect of milking hygiene to the number of bacetria in cow milk. Stočarstvo 60(6):403-411.
- Teka G (1997). Food hygiene principles and food borne disease control with special reference to Ethiopia. Faculty of Veterinary Medicine, Department of Community Health, Addis Ababa University.
- Vissers MMM, Driehuis F, Te Giffel MC, De Jong P, Lankveld JMG (2007). Short communication: quantification of the transmission of microorganisms to milk via dirt attached to the exterior of teats. J. Dairy Sci. 90(8):3579-3582.
- Wallace RL (2008). Bacteria Counts in Raw Milk.
- Walstra P, Geurts TJ, Noomen A, Van Boekel M (1999). Principles of Milk Properties and Processes. Dairy Technol. 1:149-170.
- Wafula W, Wafula JM, Masani JN (2016). Effectiveness of the Sanitation Regimes used by dairy actors to control microbial contamination of plastic jerry cans surfaces. Int. J. food Contamination 3(9):16-28.

## International Journal of Livestock Production

**Related Journals Published by Academic Journals** 

- Journal of Plant Breeding and Crop Science
- African Journal of Agricultural Research
- Journal of Horticulture and Forestry
- International Journal of Fisheries and Aquaculture
- Journal of Cereals and Oilseeds
- Journal of Soil Science and Environmental Management
- Journal of Stored Products and Postharvest Research

## academic Journals